

## **Dutch Founder SDHB Exon 3 deletion in Patients with Pheochromocytoma-Paraganglioma in South Africa**

**Debra M. Gordon <sup>1</sup>, Pablo Beckers <sup>2</sup>, Emilie Castermans <sup>2</sup>, Sebastian J.C.M.M. Neggers <sup>3</sup>, Liliya Rostomyan<sup>4</sup>, Vincent Bours <sup>2</sup>, Patrick Petrossians<sup>4</sup>, Vinciane Dideberg <sup>2</sup>, Albert Beckers <sup>4</sup>, and Adrian F. Daly <sup>4</sup>**

1. University of the Witwatersrand (WITS) Donald Gordon Medical Centre, 18 Eton Road, Parktown, Johannesburg, 2193, South Africa.

2. Department of Human Genetics, Centre Hospitalier Universitaire de Liège, Liège Université, 4000 Liège, Belgium.

3. Department of Endocrinology, Erasmus University Medical Center, Rotterdam, The Netherlands.

4. Department of Endocrinology, Centre Hospitalier Universitaire de Liège, Liège Université, 4000 Liège, Belgium.

**Short title:** Dutch founder SDHB mutation in South Africa

**Word Count:** 2689

**Keywords:** SDHB, founder mutation, paraganglioma, pheochromocytoma, South Africa, Dutch, Netherlands

### **Address for Correspondence**

Adrian F. Daly MB BCh, PhD

Department of Endocrinology,

Centre Hospitalier Universitaire de Liège,

Liège Université,

4000 Liège, Belgium

Email: [adrian.daly@uliege.be](mailto:adrian.daly@uliege.be)

## Abstract

**Objective:** Screening studies have established genetic risk profiles for diseases such as multiple endocrine neoplasia type 1 (MEN1) and pheochromocytoma-paraganglioma (PPGL). Founder effects play an important role in regional/national epidemiology of endocrine cancers, particularly PPGL. Founder effects in the Netherlands have been described for various diseases, some of which established themselves in South Africa due to Dutch emigration. The role of Dutch founder effects in South Africa have not been explored in PPGL.

**Design:** We performed a single-center study in South Africa of the germline genetic causes of isolated/syndromic neuroendocrine tumors.

**Methods:** Next-generation panel and multiplex ligand-dependent probe amplification for endocrine neoplasia risk genes.

**Results:** From a group of 13 patients we identified six with PPGL, four with sporadic or familial isolated pituitary adenomas (FIPA), and three with clinical MEN1; genetic variants were identified in 9/13 cases. We identified the Dutch founder exon 3 deletion in *SDHB* in two apparently-unrelated individuals with distinct ethnic backgrounds that had metastatic PPGL. Asymptomatic carriers with this Dutch founder *SDHB* exon 3 deletion were also identified. Other PPGL patients had variants in *SDHB*, *SDHD* and three *MEN1* variants were identified among MEN1 and young-onset pituitary adenoma patients.

**Conclusions:** This is the first identification of a Dutch founder effect for PPGL in South Africa. Awareness of the presence of this exon 3 *SDHB* deletion could promote targeted screening at a local level. Insights into PPGL genetics in South Africa could be achieved by studying existing patient databases for Dutch founder mutations in *SDHx* genes.

## Introduction

The role of germline genetic factors in neuroendocrine/endocrine neoplasias has advanced significantly in recent decades with the identification of novel genetic causes for inheritable isolated and syndromic tumors. This is particularly true in the case of pheochromocytomas and paragangliomas (PPGL) (1). These neuroendocrine tumors produce symptoms due to the synthesis and release of bioactive amines, neurotransmitters and hormones and about twenty new germline and somatic genetic factors have been discovered in recent decades (1). Among the best characterized genetic causes of PPGL are pathological variants involving genes encoding succinate dehydrogenase (SDH) subunits, *SDHA*, *SDHB*, *SDHC*, *SDHD* and *SDHAF* (collectively referred to as *SDHx*). Disruption by mutation or epimutation of *SDHx* genes leads to a state of cellular pseudohypoxia due to abnormal regulation of the Krebs cycle, accumulation of cancer inducing metabolites and subsequent activation of multiple targets of HIF1alpha. *SDHx* germline pathological variants are a major cause of familial disease, and *SDHB* accounts for about 10% of familial cases and have a relatively high risk of metastasis.

Genetic testing regimes are in place internationally for the diagnosis and characterization of genetic forms of PPGL. National datasets have identified high prevalences of particular pathological *SDHx* variants in defined geographical regions due to founder effects. For instance, endemic disease due to a p.Tyr114Cys *SDHD* variant in the Mocheni valley close to Trentino, Italy was found to originate from a common, probably Germanic founder 600-700 years previously (2). Other founder effects have been reported in *SDHx* genes in Spain, Portugal, Quebec-Canada (3, 4, 5). The strong influence of highly prevalent founder *SDHx* mutations on the national epidemiology of PPGL is well typified by the experience of the Netherlands (6). In a large national survey (n=1045 DNA samples), Hensen *et al* reported that among 690 cases/carriers with a mutation in *SDHx* genes, a full 89% had one of six Dutch founder mutations (7). In the Netherlands the *SDHD* founder mutation, c.274G>T and p.Asp92Tyr, accounted for almost 70% of all *SDHx* carriers/cases (7). *SDHB* mutations play a concomitantly lower role in the epidemiology of PPGL in the Netherlands, although *SDHB* founders have also been described there (8). In that study Bayley and colleagues reported nine apparently unrelated Dutch patients that all had an exon 3 deletion (c.201-4429\_287-933del; p.Cys68HisfsX21). The patients had extra-adrenal and head and neck paragangliomas, pheochromocytomas and pituitary adenomas; only one had a family history of PPGL (8). All nine patients shared haplotypes around *SDHB* and had identical breakpoint sequences, proving a common founder.

While founder effects can lead to an elevated regional/national prevalence of a particular *SDHx* mutation among PPGL cohorts, such individuals can “seed” new geographic foci in line with patterns of population expansion and emigration. From the late 16<sup>th</sup> century, Dutch exploration and trade led

to the establishment of settlements and colonies throughout Asia, Africa and the Americas. In South Africa, a number of genetic diseases in the population have been tied to founder effects derived from early settlers from the Netherlands (9). Whereas other founder effects are well established in South Africa and most PPGL in the Netherlands are accounted for by founder SDHx mutations, to date no Dutch founder effect has been demonstrated in PPGL patients in South Africa. Indeed, there is a general paucity of genetic risk information for PPGL and other endocrine related cancers throughout sub-Saharan Africa (10, 11, 12). As part of a two-year collaborative study, we examined the genetic causes of different endocrine and neuroendocrine tumors in a single center in South Africa. We identified multiple individuals with genetic forms of endocrine tumors, including two apparently unrelated individuals with the Dutch founder SDHB exon 3 deletion.

## Patients and Methods

The study population consisted of patients under the care of the author DMG in Johannesburg, South Africa (DG) with diagnoses of aggressive or familial neuroendocrine tumors. For inclusion patients had to have a clinical or family history consistent with the following syndromes: MEN1, MEN2, MEN4, McCune-Albright syndrome, Carney complex, familial isolated pituitary adenomas (FIPA), familial isolated hyperparathyroidism (FIHP). Patients with sporadic or syndromic PPGL, or early onset pituitary adenomas (<30 years) were also eligible for inclusion. Genetic studies were undertaken using genomic DNA from the index cases. When a pathological variant in a potential risk gene for PPGL was found, genetic testing of potential carriers in the family was also offered. Family enquiries regarding geographic origins were undertaken.

Genetic studies utilized a combination of panel based NGS and Sanger sequencing for genes of interest: MLPA was used to assess for the presence of whole or partial gene deletions. The panel of genes assessed included: *AIP*, *MEN1*, *CDKN1B*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *SDHAF2*, *VHL*, and *RET* and *MAX*. The following SALSA MLPA (MRC-Holland) were used for MLPA analyses: P244, P016, P429, P226. Next generation sequencing was amplicon-based. Briefly, all the coding exons of each gene were amplified during a first PCR with specifically designed primers in four multiplex PCRs. A second PCR was performed to incorporate the molecular identifiers and adaptors in the generated fragments. The amplicons were quantified and pooled and run on a Miseq Sequencer (Illumina).

### *Breakpoint sequencing*

For each patient presenting a deletion of *SDHB* exon 3 on the P226 MLPA, a specific PCR was performed using the following primers:

*SDHB-delE3F*: GTAATCCCAACATTCTGAGAGG

*SDHB-delE3R*: TTAAAGCCACTGTTATTTGAAC

The primers were checked for the presence of common SNPs and were blasted. The primers hybridize at intron 2 (between exon 2 and 3) and at intron 3 (between exon 3 and 4). This design allows a PCR amplification only in patients with the deletion, and generates a 262bp fragment. Sequencing was performed on an ABI 3500XL and data analysis was performed with Seqpilot 3.5.2 and Seqnext 4.3.1 (JSI Medical systems)

### *Ethics Approval*

The study was approved with the Ethics Committee of the University of Liège, Belgium and the University of the Witwatersrand, South Africa. All patients and relatives provided informed written

consent.

## Results

Assessment of clinical characteristics in the study center population identified 13 index individuals that met the inclusion criteria and underwent genetic studies (Table 1). There were six index patients with PPGL identified at the study center. Of these, two had a positive family history, the rest were clinically sporadic. In five of the six index cases a pathological genetic variant was discovered by NGS/Sanger screening: four patients had an *SDHB* pathological variant, another had an *SDHD* variant. For *SDHB*, two unrelated patients P06 and P07 carried the heterozygous c.201-4429\_287-934del (p.Cys68Hisfs\*22), a class 5 pathogenic variant. This variant leads to a large deletion of approximately 7.9 kilobases, encompassing exon 3 of *SDHB* and causes a frameshift and premature stop codon 22 amino acids downstream in exon 4. P07 had an extra-adrenal paraganglioma that recurred with metastases post operatively. The variant was not present in the mother and one child of patient P07 (two other unaffected children carried the variant); P07 had two brothers who died of PPGL previously and a sister who had been diagnosed recently with PPGL in another country. Patient P06, who has a metastatic pheochromocytoma from a young age (Figure 1), had no family members who wished to undergo screening

The *SDHB* exon 3 deletion c.201-4429\_287-934del was originally described as a Dutch founder mutation. Given the established role of Dutch founder mutations in the causation of inherited diseases in South Africa, we hypothesized that the *SDHB* exon 3 deletion might similarly be due to a founder effect. P06 and P07 shared no known family links, with P06 coming from the historical multiracial ethnic group characteristic of the Western Cape and elsewhere (14, 15). P07 is from an Afrikaner family. On discussion with the affected individuals regarding their geographical origins, no common place or person was identified and searches of church or other records for such links was not feasible due to lack of information. We sequenced the breakpoints around the *SDHB* exon 3 deletion in a known carrier of this change from the Netherlands. Thereafter we sequenced the breakpoints of the exon 3 deletion in P06 and P07. As seen in Figure 2, the exon 3 deletion breakpoints were identical in the patients from the Netherlands and South Africa, which strongly links the cases reported here to the established Dutch founder.

P13 was a clinically sporadic case that had a single neck paraganglioma, who was found to carry a c.423+1G>C probably pathological splice site variant. Interestingly, a G>A change at the same position has been described as a founder mutation in the Netherlands; this abolishes the consensus splice donor sequence and moves the normal splice site into exon 4, thereby deleting 18 amino acids (13). Patient P14 carried a c.287G>A (p.Gly96Asp) class 4 variant in *SDHB*, which was also present two nieces that were affected with paragangliomas and renal cell carcinoma; her unaffected son was a carrier. P05 who had a bilateral carotid body paraganglioma and right sided pheochromocytoma

(Figure 3) was found to have a pathological c.337\_340delGACT *SDHD* variant that leads to a predicted p.Asp113Metfs\*21 change at the protein level.

Three patients were identified that presented clinically with MEN1. Of these, two had *MEN1* variants on sequencing of *MEN1*, whereas one patient with a pituitary microadenoma, parathyroid hyperplasia and adrenal adenomas was negative for all genetic studies. The two *MEN1* variants were the truncating variant p.Thr210Serfs\*13, which has been widely identified in MEN1 cohorts, and the other was a c.824+5G>A change that is considered to be a VUS3 based on its rarity and on *in silico* prediction of affecting splicing. Among the four pituitary patients, two had FIPA, and two were sporadic. One of the sporadic patients had a large, aggressive prolactinoma at a young age (Figure 3) and was found to have a c.1618C>T; p.Pro540Ser change in *MEN1*, which has conflicting clinical significance. Among the other patients, no sequence or MLPA changes were seen, including in one patient with young onset acromegaly and a large irregular café au lait macule who also had negative digital droplet PCR for mosaic *GNAS* mutations underlying McCune Albright syndrome (16).

## Discussion

Genetic studies are playing a growing role in the clinical investigation and management of various cancers in the developed world. This is true for endocrine neoplasias like PPGL, where a large number of new target genes have been identified recently (17). In the developing world, technological and budgetary constraints conspire to limit greatly the availability and use of clinical genetics, thereby hindering optimal management. For example, in sub-Saharan Africa, there is significant and growing cancer burden but a relative dearth of information about underlying genetic risk profiles among highly heterogeneous populations (18). In South Africa there is the added complexity of large numbers of individuals with European, Asian and mixed heritage in addition to different Black populations. Therefore, the identification of existing genetic risks in European populations and elsewhere is relevant in South Africa. For endocrine neoplasias in particular, the identification of potentially inheritable PPGL risk genes is of clear clinical importance as large-scale screening in Europe has revealed enrichment of certain mutations (*SDHx*) among subpopulations (geographic, religious, cultural, ethnic), leading to endemic PPGL genetic risk profiles (2, 7). When carriers emigrate from areas of endemic genetic disease to new regions they can act as founders of new foci of genetic risk.

In this study we found a new Dutch founder effect in South Africa, due to the presence of a characteristic *SDHB* exon 3 deletion causing familial and sporadic PPGL. This exon 3 deletion in *SDHB* was first described in the Netherlands by Bayley *et al* (8). In that study they described nine apparently unrelated Dutch patients with sporadic PPGL and performed breakpoint analysis at the deletion; all nine had the same sequencing findings. Using an identical approach, we demonstrated a 1.6 kB PCR product in the two South African PPGL cases and in a known Dutch carrier of the exon 3 *SDHB* deletion. Sequencing of the breakpoints in the South African cases and in the Dutch case was identical. This finding is unlikely to have occurred due to a mutational hotspot. As noted by Bayley *et al*, the downstream breakpoint is not located in an Alu repeat region and there are no other features that would make this a likely region for a hotspot (8).

Previous studies in the Netherlands and among populations of Dutch origin worldwide provide some of the best examples of the importance of founder effects in genetic diseases. The Netherlands exemplifies the role of geographic and cultural factors in the establishment and subsequent enrichment of pathological genetic variants in their population (9). Study of Dutch founder mutations has shown that some arose within culturally, geographically and family-determined genetic isolates within the Netherlands. As religious and cultural limitations loosened in the 20<sup>th</sup> century and the Dutch population increased, these founder mutations came to play an important role nationally in genetic disease epidemiology (9). PPGL genetics is a good example of this. The overwhelming

majority of PPGL in the Netherlands are derived from specific *SDHD* mutations, particularly c.274G>T and c.416T>C (6). Indeed, nearly 89% of all *SDHx* carriers in the Netherlands have one of six founder mutations in *SDHD*, *SDHB* and *SDHAF*; the *SDHB* Exon 3del is one of these (6, 7, 19). Emigration of Dutch settlers to colonies in South Africa, the Americas and elsewhere led to the establishment of new concentrations of carriers of founder mutations in various genes. In South Africa, numerous well described examples of Dutch founder effects in diseases related to hypercholesterolemia, porphyria have even revealed the likely identity of the founder centuries before. The current study strongly suggests that an *SDHB* founder mutation might similarly be playing a role in PPGL pathophysiology in South Africa.

The penetrance of *SDHB* mutation related PPGL is variable (25-75%); data from large kindreds suggest that even the low end of this range may well be a significant over-estimate (20). This low penetrance would explain the lack of a recognized family history of PPGL in patient P06, which echoes the sporadic presentation of the nine Dutch PPGL patients in the original report of the *SDHB* exon 3 del founder (21). Identification of this founder effect in South Africa is clinically relevant, as the patients and families affected in this study had an aggressive phenotype. Further study of previously identified patients with PPGL in South Africa could provide more information on the relative frequency of the exon 3 deletion *SDHB* founder mutation (10, 22, 23).

Other genetic causes of PPGL in the current study include a truncating *SDHD* c.337\_340delGACT (p.Asp113Metfs\*21) change in a sporadic male of Indian origin with bilateral head and neck paraganglioma and a pheochromocytoma. This pathological variant had been identified in various sporadic and familial PPGL cohorts from Asturias (Spain), India and the Netherlands (7, 24, 25). Another patient with a European Jewish background had a family history of PPGL in two nieces and all three shared the same c.287G>A (p.Gly96Asp) *SDHB* change; this missense change was previously reported in the United States and elsewhere (26, 27, 28). Among the patients with presentations typical of MEN1, in two cases previously identified *MEN1* variants were identified in germline DNA. In the patients with FIPA or isolated aggressive pituitary adenomas, no variants in *AIP* were seen, which is in keeping with the known genetic epidemiology of these populations (29). One aggressive prolactinoma was noted in a young patient who had a variant of unknown significance in *MEN1*; there is a known association between *MEN1* variants and young onset pituitary adenomas (30).

The study has a number of limitations. As a small single center cohort, we cannot determine the overall structure of the genetic risk profile for endocrine neoplasias, including PPGL in South Africa as a whole, similar to previous genetic studies on this topic in South Africa (10, 11, 12, 23). Hence, it remains to be proven in larger series whether the *SDHB* Exon 3 del mutation plays a major role in PPGL risk nationally in South Africa. Also, the limited study size means that the potential presence

in South Africa of other important Dutch founder *SDHx* mutations, particularly *SDHD* c.274G>T, remains to be established. Similarly, the presence of *SDHB* Exon 3 del in other Dutch emigrant populations will need to be specifically addressed.

In conclusion, this two-year study of endocrine neoplasia population at single center in South Africa identified multiple patients with MEN, PPGL and pituitary adenomas with clinically-actionable genetic variants. In particular, we identified for the first time the presence of the known Dutch founder *SDHB* Exon 3 del mutation among apparently unrelated PPGL patients in South Africa. This extends the acknowledged role of Dutch founder mutations in disease into the field of inherited neuroendocrine tumors, including pheochromocytomas and paragangliomas. Wider screening programs of PPGL patients in South Africa could help to ascertain the relative importance of this and potentially other *SDHx* gene founder mutations derived from historical Dutch emigration.

**Disclosure:** The authors have nothing to disclose

**Funding:** The study was supported in part by a grant from the Fonds pour la recherche scientifique (FIRS), CHU de Liege 2017-2020

**Acknowledgements:** The authors would like to acknowledge Dr. Carli MJ Tops, Dr. Jean-Pierre Bayley, and Dr. Frederik Hes of the Departments of Clinical Genetics and Human Genetics, Leiden University Medical Center for their valuable guidance regarding Dutch founder *SDHB* mutations and Dr Manuel Van Den Venter from Lancet Laboratory in South Africa and Dr June Fabian from the research dept of WITS Donald Gordon Medical Centre for assistance in conducting the study analyses.

## Legends

**Table 1.** Clinical details of South African patients with endocrine neoplasia.

**Figure 1.** Imaging and pathology images in patient P06 with the Dutch founder exon 3 deletion of *SDHB*. CT (A) and PET-CT images showing metastatic deposits of pheochromocytoma in the pelvic and iliac regions. High-power (C) and low-power (D) H&E staining images of metastatic pheochromocytoma deposits in the iliac and peri-vesicular lymph nodes at presentation.

**Figure 2.** Sequences and MLPA results in two South African individuals with the Dutch founder exon 3 deletion in *SDHB* (Panel A, B). The identical control sequence from an established carrier of the Dutch founder exon 3 *SDHB* deletion is shown in Panel C.

**Figure 3.** Panel A shows CT of patient P05 with bilateral carotid body paraganglioma due to a pathogenic germline *SDHD* variant (c.337\_340delGACT; p.Asp113fs). Panel B shows a coronal MRI image at diagnosis of a sporadic pituitary macroadenoma (prolactinoma) in patient P05 with a *MEN1* variant (c.1618C>T; p.Pro540Ser).

## References

1. Papathomas TG, Suurd DPD, Pacak K, Tischler AS, Vriens MR, Lam AK, & Krijger RR de. What Have We Learned from Molecular Biology of Parangliomas and Pheochromocytomas? *Endocrine Pathology* 2021. pp 134–153. . (doi:10.1007/s12022-020-09658-7)
2. Schiavi F, Dematte S, Cecchini ME, Taschin E, Bobisse S, Piano A Del, Donner D, Barbareschi M, Manera V, Zovato S, et al. The endemic paraganglioma syndrome type 1: origin, spread, and clinical expression. *J Clin Endocrinol Metab* 2012 **97** E637-41. (doi:10.1210/jc.2011-2597)
3. Bourdeau I, Grunenwald S, Burnichon N, Khalifa E, Dumas N, Binet MC, Nolet S, & Gimenez-Roqueplo AP. A SDHC founder mutation causes paragangliomas (PGLs) in the French Canadians: New insights on the SDHC-related PGL. *Journal of Clinical Endocrinology and Metabolism* 2016 **101** 4710–4718. (doi:10.1210/jc.2016-1665)
4. Cascón A, Pita G, Burnichon N, Landa I, López-Jiménez E, Montero-Conde C, Leskelä S, Leandro-García LJ, Letón R, Rodríguez-Antona C, et al. Genetics of pheochromocytoma and paraganglioma in Spanish patients. *Journal of Clinical Endocrinology and Metabolism* 2009 **94** 1701–1705. (doi:10.1210/jc.2008-2756)
5. Martins RG, Nunes JB, Máximo V, Soares P, Peixoto J, Catarino T, Rito T, Soares P, Pereira L, Sobrinho-Simões M, et al. A founder SDHB mutation in Portuguese paraganglioma patients. *Endocrine-Related Cancer* 2013. (doi:10.1530/ERC-12-0399)
6. Hensen EF, Siemers MD, Jansen JC, Corssmit EPM, Romijn JA, Tops CMJ, Mey AGL Van Der, Devilee P, Cornelisse CJ, et al. Mutations in SDHD are the major determinants of the clinical characteristics of Dutch head and neck paraganglioma patients. *Clinical Endocrinology* 2011 **75** 650–655. (doi:10.1111/j.1365-2265.2011.04097.x)
7. Hensen EF, Duinen N van, Jansen JC, Corssmit EPM, Tops CMJ, Romijn JA, Vriends AHJT, Mey AGL van der, Cornelisse CJ, Devilee P, et al. High prevalence of founder mutations of the succinate dehydrogenase genes in the Netherlands. *Clinical Genetics* 2012 **81** 284–288. (doi:10.1111/j.1399-0004.2011.01653.x)
8. Bayley JP, Grimbergen AEM, Bunderen PA van, Wielen M van der, Kunst HP, Lenders JW, Jansen JC, Dullaart RPF, Devilee P, Corssmit EP, et al. The first Dutch SDHB founder deletion in paraganglioma - Pheochromocytoma patients. *BMC Medical Genetics* 2009 **10** . (doi:10.1186/1471-2350-10-34)

9. Zeegers MPA, Poppel F van, Vlietinck R, Spruijt L, & Ostrer H. Founder mutations among the Dutch. *European Journal of Human Genetics* 2004 **12** 591–600. (doi:10.1038/sj.ejhg.5201151)
10. Bruce-Brand C & Wyk AC Van. Prevalence of succinate dehydrogenase deficiency in paragangliomas and pheochromocytomas at a tertiary hospital in Cape Town: a retrospective review. *Journal of Endocrinology, Metabolism and Diabetes of South Africa* 2021 **26** 9–15. (doi:10.1080/16089677.2020.1838161)
11. Shone D, Goedhals J, & Pearce NE. Malignant paraganglioma in an African patient associated with a succinate dehydrogenase subunit B (SDHB) mutation. *South African Journal of Surgery* 2018 **56** 64–66. (doi:10.17159/2078-5151/2018/V56N2A2391)
12. Siddiqui N, Seedat F, Bulbulia S, Mtshali NZ, Botha A, Krause A, Daya R, & Bayat Z. SDHB-Associated Paraganglioma Syndrome in Africa-A Need for Greater Genetic Testing. *Journal of the Endocrine Society* 2021 **5** 1–6. (doi:10.1210/jendso/bvab111)
13. Bayley JP, Minderhout I van, Weiss MM, Jansen JC, Oomen PHN, Menko FH, Pasini B, Ferrando B, Wong N, Alpert LC, et al. Mutation analysis of SDHB and SDHC: Novel germline mutations in sporadic head and neck paraganglioma and familial paraganglioma and/or pheochromocytoma. *BMC Medical Genetics* 2006 **7** . (doi:10.1186/1471-2350-7-1)
14. Quintana-Murci L, Harmant C, Quach H, Balanovsky O, Zaporozhchenko V, Bormans C, Helden PD van, Hoal EG, & Behar DM. Strong Maternal Khoisan Contribution to the South African Coloured Population: A Case of Gender-Biased Admixture. *American Journal of Human Genetics* 2010 **86** 611–620. (doi:10.1016/j.ajhg.2010.02.014)
15. Statistics South Africa. Mid year population estimates 2019 (STATISTICAL RELEASE P0302). 2019.
16. Vasilev V, Daly AF, Thiry A, Petrossians P, Fina F, Rostomyan L, Silvy M, Enjalbert A, Barlier A, & Beckers A. McCune-Albright syndrome: a detailed pathological and genetic analysis of disease effects in an adult patient. *The Journal of clinical endocrinology and metabolism* 2014 **99** E2029-38. (doi:10.1210/jc.2014-1291)
17. Juhlin CC. Challenges in Paragangliomas and Pheochromocytomas: from Histology to Molecular Immunohistochemistry. *Endocrine Pathology* 2021. pp 228–244. . (doi:10.1007/s12022-021-09675-0)
18. Rotimi SO, Rotimi OA, & Salhia B. A Review of Cancer Genetics and Genomics Studies in Africa. *Frontiers in Oncology* 2020 **10** 1. (doi:10.3389/FONC.2020.606400)
19. Taschner PEM, Jansen JC, Baysal BE, Bosch A, Rosenberg EH, Bröcker-Vriends AHJT, Mey

- AGL Van Der, Ommen GJB Van, Cornelisse CJ, & Devilee P. Nearly all hereditary paragangliomas in the Netherlands are caused by two founder mutations in the SDHD gene. *Genes Chromosomes and Cancer* 2001 **31** 274–281. (doi:10.1002/gcc.1144)
20. Rijken JA, Niemeijer ND, Corssmit EPM, Jonker MA, Leemans CR, Menko FH, & Hensen EF. Low penetrance of paraganglioma and pheochromocytoma in an extended kindred with a germline SDHB exon 3 deletion. *Clinical Genetics* 2016 **89** 128–132. (doi:10.1111/cge.12591)
  21. Bayley JP, Grimbergen AE, Bunderen PA van, Wielen M van der, Kunst HP, Lenders JW, Jansen JC, Dullaart RP, Devilee P, Corssmit EP, et al. The first Dutch SDHB founder deletion in paraganglioma-pheochromocytoma patients. 2009 . (doi:10.1186/1471-2350-10-34)
  22. Huddle K. Pheochromocytoma in black South Africans - a 30-year audit. *South African medical journal Suid-Afrikaanse tydskrif vir geneeskunde* 2011 **101** 184–188. (doi:10.7196/SAMJ.4320)
  23. Nel D, Panieri E, Malherbe F, Steyn R, & Cairncross L. Surgery for Pheochromocytoma: A Single-Center Review of 60 Cases from South Africa. *World Journal of Surgery* 2020 **44** 1918–1924. (doi:10.1007/s00268-020-05420-6)
  24. Lima J, Feijão T, Silva AF Da, Pereira-Castro I, Fernandez-Ballester G, Máximo V, Herrero A, Serrano L, Sobrinho-Simões M, & Garcia-Rostan G. High frequency of germline succinate dehydrogenase mutations in sporadic cervical paragangliomas in Northern Spain: Mitochondrial succinate dehydrogenase structure-function relationships and clinical-pathological correlations. *Journal of Clinical Endocrinology and Metabolism* 2007 **92** 4853–4864. (doi:10.1210/jc.2007-0640)
  25. Kapoor N, Pai R, Ebenazer A, Sen I, Stephen E, Agarwal S, Paul MJ, & Rajaratnam S. Familial carotid body tumors in patients with sdhd mutations: A case series. *Endocrine Practice* 2012 **18** . (doi:10.4158/EP12012.CR)
  26. Brouwers FM, Eisenhofer G, Tao JJ, Kant JA, Adams KT, Linehan WM, & Pacak K. High frequency of SDHB germline mutations in patients with malignant catecholamine-producing paragangliomas: Implications for genetic testing. *Journal of Clinical Endocrinology and Metabolism* 2006 **91** 4505–4509. (doi:10.1210/jc.2006-0423)
  27. Ghayee HK, Havekes B, Corssmit EPM, Eisenhofer G, Hammes SR, Ahmad Z, Tessnow A, Lazúrová I, Adams KT, Fojo AT, et al. Mediastinal paragangliomas: association with mutations in the succinate dehydrogenase genes and aggressive behavior. *Endocrine-Related Cancer* 2009 **16** 291–299. (doi:10.1677/ERC-08-0214)
  28. Benn DE, Gimenez-Roqueplo AP, Reilly JR, Bertherat J, Burgess J, Byth K, Croxson M,

- Dahia PL, Elston M, Gimm O, et al. Clinical presentation and penetrance of pheochromocytoma/paraganglioma syndromes. *J Clin Endocrinol Metab* 2006 **91** 827–836. (doi:10.1210/jc.2005-1862)
29. Vandeva S, Daly AF, Petrossians P, Zacharieva S, & Beckers A. Somatic and germline mutations in the pathogenesis of pituitary adenomas. *European Journal of Endocrinology* 2019 **181** . (doi:10.1530/EJE-19-0602)
30. Cuny T, Pertuit M, Sahnoun-Fathallah M, Daly A, Occhi G, Odou MF, Tabarin A, Nunes ML, Delemer B, Rohmer V, et al. Genetic analysis in young patients with sporadic pituitary macroadenomas: besides AIP don't forget MEN1 genetic analysis. *European journal of endocrinology / European Federation of Endocrine Societies* 2013 **168** 533–541. (doi:10.1530/EJE-12-0763)

Patient Number	Sex	Age (years)	Tumors	Family cancer history	Genetic result
<b>MEN Syndromes</b>					
P01	F	48	Adrenal adenomas, pituitary microadenoma, parathyroid hyperplasia (2 glands)	Colorectal carcinoma, astrocytoma, liposarcoma	No pathological variants on panel or MLPA
P02	F	23	Pituitary macroadenoma (prolactinoma); Parathyroid hyperplasia (3 glands); Multiple pancreatic NET	None	<b>MEN1</b> c.628_631delACAG p.Thr210Serfs*13
P03	M	40	Hyperparathyroidism (multiple gland)	Mother with clinical MEN1	<b>MEN1</b> c.824+5G>A VUS3
<b>PPGL</b>					
P05	M	33	Bilateral carotid body paraganglioma, right sided pheochromocytoma	None	<b>SDHD</b> c.337_340delGACT p.Asp113fs
P06	F	16	Metastatic malignant pheochromocytoma	None	<b>SDHB</b> c.201-4429_287-934del (p.Cys68Hisfs*22 Exon 3 deletion
P07	F	35	Metastatic paraganglioma	Two brothers died from pheochromocytoma, one sister PPGL. Two unaffected children are carriers.	<b>SDHB</b> c.201-4429_287-934del (p.Cys68Hisfs*22 Exon 3 deletion
P12	F	16	Pheochromocytoma	None	No pathological variants on panel or MLPA
P13	F	40	Neck paraganglioma	None	<b>SDHB</b> c.423+1G>C Splice site pathological variant
P14	F	45	Cardiac paraganglioma	Two nieces with paraganglioma, paraganglioma +renal cell cancer. One unaffected child is a carrier.	<b>SDHB</b> c.287G>A p.Gly96Asp
<b>Pituitary</b>					

P04	F	28	Pituitary macroadenoma (prolactinoma), resistant to dopamine agonists; neurosurgery (x2)	None	<b>MEN1</b> c.1618C>T p.Pro540Ser
P21	M	25	Acromegaly (macroadenoma, resistant to somatostatin analogs); café au lait macule	None	No pathological variants on panel or MLPA. Digital Droplet PCR negative for <i>GNAS</i> mosaic mutations
P22	F	Mid-teens	Prolactinoma (macroadenoma) that responded to cabergoline; in early 20's developed acromegaly (microadenoma)	FIPA (first cousin macroprolactinoma)	No pathological variants on panel or MLPA
P23	F	20's	Prolactinoma (microadenoma); low bone mineral density; dysembryoplastic neuroepithelial tumor	FIPA (unknown adenoma subtype)	No pathological variants on panel or MLPA

Table 1. Characteristics of the study population.

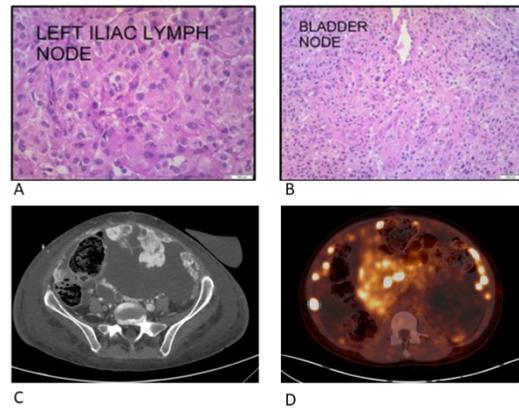


Figure 1

Figure 1

338x190mm (96 x 96 DPI)

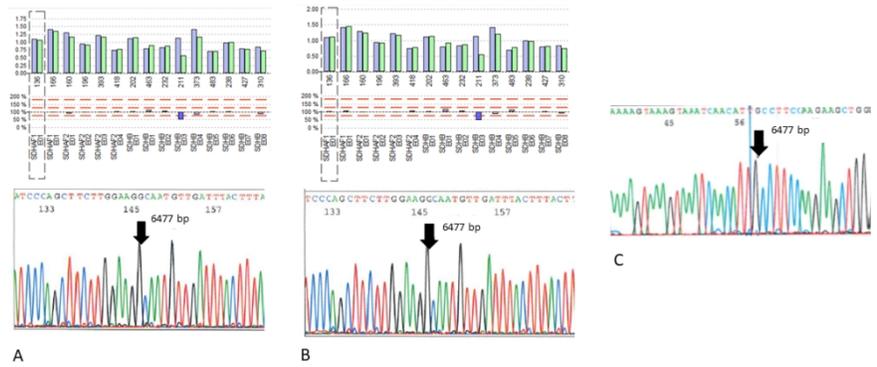


Figure 2

Figure 2

338x190mm (96 x 96 DPI)



Figure 3

Figure 3

338x190mm (96 x 96 DPI)